cerabone®
Natural bovine bone grafting material
Scientific and clinical evidence
Bone and regeneration techniques

The use of bone graft materials

Bone graft materials are applied to replace and regenerate bone matrix lost by various reasons such as tooth extraction, cystectomy or bone atrophy following loss of teeth or inflammatory processes. For the filling of bone defects, the patient’s own (autologous) bone is considered the “gold standard”, because of its biological activity due to vital cells and growth factors. Nevertheless, the harvesting of autologous bone requires a second surgical site associated with an additional bony defect and potential donor site morbidity.

In addition, the quantity of autologous bone is limited. Today, due to a constant development, bone graft materials provide a reliable and safe alternative to autologous bone grafts. Clinicians can choose between a variety of different bone graft materials and augmentation techniques. Bone graft materials are classified by their origin into four groups.

The GBR/GTR technique

The principle of Guided Bone Regeneration (GBR) or Guided Tissue Regeneration (GTR) is based on the separation of the grafted site from the surrounding soft tissue by application of a barrier. Collagen membranes act as a resorbable matrix to avoid the ingrowth of the faster proliferating fibroblasts and/or epithelium into the defect, and to maintain the space for controlled regeneration of bone. The application of a bone graft material into the defect prevents a collapse of the collagen membrane, acting as a place holder for the regenerating bone and as an osteoconductive scaffold for the ingrowth of blood vessels and bone forming cells.

Classification

Autologous:
- Patient’s own bone, mostly harvested intraorally or from the iliac crest
- Intrinsic biological activity

Allogenic:
- Bone from human donors
- Natural bone composition and structure

Xenogenic:
- From other organisms, mainly bovine origin
- Long-term volume stability

Alloplastic:
- Made of synthetic materials
- No risk of disease transmission

For large defects a mixture of autologous or allogenic bone, which has excellent biological potential, and a bone graft material for volume stability of the grafting site, is recommended.
Xenogenic bone graft materials

Xenogenic bone grafts are derived from animals, preferably of bovine origin. Bovine bone materials are deproteinized by heating (sintering) to minimize the risk of allergic reactions and disease transmission. Bovine bone materials have a long tradition, are very well documented, and their clinical application has found wide-ranging acceptance. The removal of all proteins transforms them into biologically derived hydroxyapatite ceramics. They are characterized by their preserved three-dimensional natural bone structure with interconnecting pores, strongly resembling the human bone structure. Their guided osseous integration rather than rapid resorption leads to excellent volume stability of the graft, with the formation of new bone on the highly structured bovine bone surface.

cerabone® – natural bovine bone grafting material

cerabone® is derived from bovine bone in an established high-temperature heating process (sintering) guaranteeing high safety. Beside safety and reliability of the product and the production process, the material fulfills all other important requirements for the clinical success of a bovine bone graft material:

- Phase pure hydroxyapatite without organic components
- Rough and open porous structure comparable to native human bone
- Excellent hydrophilicity enabling a rapid uptake of blood
- Optimal biocompatibility proven in various in vitro and in vivo tests
- Rapid and controlled osseous integration

These characteristics are the base for the excellent clinical results of cerabone® demonstrated by high volume stability at the graft site, complete integration into newly formed bone matrix with high bone density.

Indications for cerabone®

Periodontology

- Intraosseous defects (1 - 3 walls)
- Furcation defects (class I - II)

Implantology and Oral and CMF Surgery

- Sinus floor elevation
- Horizontal augmentation
- Vertical augmentation
- Ridge preservation
- Peri-implant defects
- Socket preservation
- Bone defect augmentation

Product Specifications

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cerabone®: Safety and reliability facts
made in Germany

Patented manufacturing process
Both, product and process of procurement of the raw material as well as the production process are fulfilling the German and EU-regulatory and security requirements for bovine bone grafts including EN ISO 22442-1, -2 and -3, as well as Commission Regulation (EU) No 722/2012. The proprietary manufacturing process of cerabone® is based on high-temperature heating (sintering) and special surface treatment that result in:
- Cell-friendly, biomimetically structured, rough surface
- Complete removal of organic components and albuminous impurities
- No risk of allergic reactions or rejection

CE marking
- CE certification of cerabone® was issued in 2002
- The product is on the market since January 2002

Sterile and storable
Cerabone® is available as granules and in block form. The product is packed in sterile vials, sealed in primary and secondary blister packaging and sterilized with gamma irradiation. Cerabone® can be stored at room temperature for up to three years.

X-ray diffractometry: mineral phases and crystallinity.
Narrow peaks and low baseline.
Cerabone® shows high crystallinity and 100% purity.

Infrared spectroscopy: molecular fingerprint.
Characteristic peaks of phosphate and hydroxy groups of the hydroxyapatite.
No other organic phases detectable.

Thermogravimetric analysis showing combustion of organic components.
No mass loss by heating cerabone® up to 1000°C.
Complete removal of organic components (cells, collagen) by sintering process.

cerabone® consists of the pure mineral phase of bovine bone.
No other phases besides hydroxyapatite are detectable. The high phase purity leads to maximal volume stability. In addition, the absence of organic components ensures the high safety of cerabone®.

Prof. Dr. C. Vogt, University of Hannover

cerabone® serves as an excellent matrix for bone regeneration

Growth of osteoblasts and osteoclasts on cerabone®

In vitro results from Prof. Dr. H. Jennissen and Dr. M. Laub
University of Duisburg-Essen/Morphoplant GmbH

The rough surface also promotes the adhesion of serum proteins and cells onto the surface. Osteoblast-like cells quickly adhere to the cerabone® particles. Only attached osteoblasts can start to produce new bone matrix leading to the osseous integration of the cerabone® particles. In another study, good adherence of osteoclasts promoted the superficial remodeling of the particles.

Proliferation of osteoblasts on cerabone®

day 0           day 3          day 7
600
400
200
0
cells/well

Bound BMP-2 (mg/g cerabone®)

BMP-2 structure

Colonization of cerabone® by osteoblasts
Prof. Dr. D. Rothamel, University of Bonn

Hydrophilicity of cerabone®

cerabone®’s rapid and complete hydration with blood or saline solution is crucial for superior handling characteristics, new bone formation and for the final clinical success.

Excellent hydrophilicity of cerabone®

Its strong capillary action facilitates fast and efficient penetration of the material particles with fluids, nutrients and blood through the three-dimensional, porous trabecular bone network, resulting in excellent handling, reliability and predictability in the daily clinical use.

Scanning electron microscope (SEM) pictures show the highly structured surface of cerabone® as well as the macro and micropores.

The macroporous structure enables migration of cells, penetration of blood vessels and integration of the particles.

The capillary effect of the micropores leads to a quick blood uptake of the material.

The rough surface ensures an excellent and homogenous surface adhesion of cells and proteins.

Dr. C. Reichert, University of Bonn

Growth of osteoblasts and osteoclasts on cerabone®

In vitro results from Prof. Dr. D. Rothamel, University of Düsseldorf and Dr. C. Reichert, University of Bonn

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Bound BMP-2 (mg/g cerabone®)

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Optimal adhesion and ingrowth of cells, proteins and blood vessels

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1 Seidel and Dingeldein (2004). Cerabone® - Bovine Based Spongiosa


Stem cell research

Interaction of cerabone® with stem cells

In vitro results from Prof. Dr. B. Zavan, University of Padova

Cerabone® supports the differentiation of attached stem cells into osteoblasts that produce new bone matrix.

Collagen, osteopontin, osteonectin and osteocalcin are proteins of the extracellular bone matrix that can be used as markers for bone formation. Their detection 14 days after seeding stem cells on cerabone® indicate the correct differentiation of the cells.

Collagen I osteopontin osteonectin osteocalcin

Tissue integration and cellular degradation

In vivo data from a mouse model by Dr. Dr. S. Ghanaati, University of Mainz and University of Frankfurt a. M.

15 days after implantation into the subcutaneous tissue (CT) of mice, cerabone® (CB) is embedded within a well vascularized granulation tissue (blood vessels marked by arrows). No fibrous encapsulation or inflammatory reactions are observed. Mononuclear and multinuclear cells (arrow heads) indicate the onset of cellular degradation of the cerabone® particles.

Maximal stability and good osseous integration of cerabone®

Histological studies on cerabone®

Optimal bone regeneration after bone defect treatment with cerabone® was demonstrated in an animal study.

Bony defects following apicoectomy, were filled with cerabone®.

The histological examination showed a complete bridging of the osteotomy orifice after three months and a well established new bone (NB) and cementum formation (CEM) around the cerabone® particles.

Endodontics

Cerabone® – osteoconduction and bony regeneration

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Implantology

Cerabone® – osseous integration and optimal stability

Sinus lift study from Prof. Dr. D. Rothamel, University of Düsseldorf

A study on 12 patients showed that cerabone® acts as an osteoconductive material that supports the regeneration of bone after sinus floor elevation surgery. After six months the particles of all biopsies were completely integrated into the newly formed bone matrix, while the grafted area showed excellent volume stability.

Stem cells seeded on cerabone®

Collagen, osteopontin, osteonectin and osteocalcin

Results from Prof. Dr. Z. Artzi, University of Tel Aviv


In vivo

Compressive force (N) 1670±120 4510±770

Compressive resistance (N/cm²) 420±32 564±96

Shear force (N/cm²) 124±35 338±200

Dimension (mm) 20 20

20 20

20 40

40

5

5

4

3

2

1

optical density

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Clinical application of cerabone®

Clinical case by
Dr. Marius Steigmann, Neckargemünd, Germany

cerabone® for coverage of implant dehiscence and ridge augmentation

Extraction of tooth 21 after endodontic treatment
Application of collacone® for stabilization of the blood clot
Buccal bone defect after eight weeks healing time
A periodontal probe demonstrates the vertical extension of the defect

Implant placed into the former extraction socket
Surface of the implant is covered with autologous bone
Coverage of the autologous bone with cerabone® (0.5 - 1.0 mm)

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Surface of the implant is covered with autologous bone
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Coverage of the bone substitute with Jason® membrane
Closure of the site using single sutures after peristomeum slitting
Tension-free suturing maintains undisturbed healing
Abutment installation after implant uncovering, six months after implantation
Final prosthetic restoration with a full-ceramic crown

Contour maintenance
For augmentations in the aesthetic region cerabone® provides long-term dimensional stability and therefore a good bone bed to support an optimal contour of the soft tissue and sustained aesthetic result.

Clinical application of cerabone®

Clinical case by
Dr. Viktor Kalenchuk, Chernivtsi, Ukraine

Ridge augmentation with cerabone® and collprotect® membrane

cerabone® particles size 0.5 - 1.0 mm in place
Covering augmentation site with collprotect® membrane

Clinical situation with narrow alveolar ridge in the lower jaw
3.5 mm dental implants inserted with inefficient immersion of dental implant platforms
Implants inserted and cortical bone perforated, vestibular view

Impants uncovered, good integration of cerabone® particles
Alveolar ridge form and size renewal around implants with cerabone®

Situation at re-entry six months post-operative, implants partly covered by new bone matrix

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For augmentations in the aesthetic region cerabone® provides long-term dimensional stability and therefore a good bone bed to support an optimal contour of the soft tissue and sustained aesthetic result.

Particle Size
Small cerabone® particles (0.5 - 1.0 mm) allow a good adaptation to surface contours; they are especially useful for lateral augmentations or to fill voids when working with autologous bone blocks.
For sinus lift and extensive augmentations the use of cerabone® particle size 1.0 - 2.0 mm is recommended. The increased space between the large particles enables a better vascularization and improves the regeneration of larger defects.

Rehydration
Due to its excellent hydrophilicity, cerabone® particles adhere to each other after mixing with blood or sterile saline solution, allowing optimal handling and good adaptation to surface contours.
Clinical application of cerabone®

**Clinical case by**

Dr. Marius Steigmann, Neckargemünd, Germany

cerabone® for horizontal augmentation

- Atrophic alveolar ridge in the left mandible
- After mucoperiosteal flap elevation, the extensive bone resorption is visible
- Clinical view six months after augmentation reveals healthy soft tissue situation
- Pre-operative cone beam scan revealing good osseous formation of the augmented site
- The wide ridge allows for stable insertion of the two implants
- Situation after healing of the soft tissue
- Excellent bone regeneration six months after application of cerabone® particles and Jason® membrane
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- Excellent bone regeneration six months after application of cerabone® particles and Jason® membrane
- Insertion of gingiva formers allow for soft tissue maturation
- Final prosthetic restoration with ceramic bridge

**Antibiotic prophylaxis**

Make sure that the patient’s blood contains a sufficient concentration of antibiotics before starting the augmentation (especially for larger augmentation volumes), e.g. by starting the antibiotic one day prior to surgery or at least one hour before by ingestion of a full daily dose.

Clinical application of cerabone®

**Clinical case by**

Dr. Derk Siebers, Berlin, Germany

Socket management/ridge preservation with cerabone® and Jason® membrane

- Pre-operative OPG, tooth 25 planned for extraction
- Large defect of the buccal wall visible after tooth extraction
- Jason® membrane placed to close the mouth-antrum connection
- Socket filled with cerabone® and augmentation of the buccal wall
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- Jason® membrane placed to close the mouth-antrum connection
- Socket filled with cerabone® and augmentation of the buccal wall
- Covering of augmentation site with Jason® membrane
- Augmentation site covered with Jason® membrane in double layer
- Jason® membrane protecting the socket
- Jason® fleece placed over membrane
- Jason® fleece left exposed for open healing
- Situation two weeks after healing
- Soft tissue situation after four months healing time
- Implant insertion 12 months after augmentation
- Final prosthetic restoration, occlusal view
- Final prosthetic restoration, vestibular view

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- Final prosthetic restoration, vestibular view
Two-stage sinus lift with cerabone® and Jason® membrane

Pre-operative OPG
Preparation of a lateral window
Perforation of the Schneiderian membrane visible after preparation of the lateral window
Jason® fleece introduced into the sinus cavity to cover the Schneiderian membrane
Filling of the sinus cavity with cerabone®
Simultaneous placement of three implants
Additional horizontal augmentation with cerabone®
Covering of the augmentation site with Jason® membrane
Re-opening six months after implantation, stable integration of the cerabone® particles
Placement of gingiva formers
Good situation after removal of gingiva formers, six weeks after re-opening

Schneiderian membrane perforation
In case of a small perforation (< 5 mm) of the Schneiderian membrane during sinus floor elevation, the application of a collagen membrane (e.g. Jason® membrane or collprotect® membrane) is a useful tool for perforation coverage. Instruct the patient to avoid sneezing for two weeks and prescribe antibiotics and swelling prophylaxis (e.g. xylometazoline). Never continue if you find an acute sinusitis with presence of pus.

Membrane coverage
For better and more predictable results we always recommend to cover the augmentation area (and the lateral sinus window after sinus floor elevation) with a collagen membrane (e.g. collprotect® membrane or Jason® membrane).
Clinical application of cerabone®

Clinical case by
Dr. Damir Jelušić, Opatija, Croatia

Socket preservation with cerabone®

Pre-operative CT of teeth eleven and 21 after endodontic treatment

Teeth eleven and 21 not worth saving and planned for extraction

Situation after extraction of the front teeth

Jason® membranes placed within in the extraction sockets, covering the vestibular wall

Filling of the sockets with cerabone®

Jason® membrane turned down over the socket and sutured

Post-operative CT four months after extraction, good preservation of the ridge

Placement of gingiva formers

Final prosthetic situation with individual emergence profile created with provisional crowns (four months post implantation)

Individualized zirconium abutments

Final prosthetic restoration with ceramic crowns

Density
Avoid to compress the cerabone® particles excessively at the defect site. Open space between the particles permits blood vessel ingrowth and the formation of new bone matrix.

Clinical case by
Dr. Raluca Cosgarea and Prof. Dr. Dr. Anton Sculean, Cluj-Napoca, Romania and Bern, Switzerland

Regeneration of intrabony defects with cerabone® and collprotect® membrane

Pre-operative defect measurement

Pre-operative x-ray showing intrabony defect

Defect presentation after preparation of mucoperiosteal flap

Rehydration of cerabone® particles

collprotect® membrane cut to shape

Filling of intrabony defect with cerabone®

collprotect® membrane in place

Wound closure

X-ray control at 12 months post-operative

X-ray at 24 months post-operative

Final prosthetic restoration

Sterile application
Pay attention to sterile application of the substitute, e.g. by using new instruments for granule insertion (and trimming of membranes). Prior contact to saliva may contaminate your graft.
Innovation.
Regeneration.
Aesthetics.

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